

CrossMark

Urine TMPRSS2: ERG Fusion Transcript as a Biomarker for Prostate Cancer: Literature Review

Francesca Sanguedolce,¹ Antonella Cormio,² Matteo Brunelli,³ Alessandro D'Amuri,⁴ Giuseppe Carrieri,⁵ Pantaleo Bufo,¹ Luigi Cormio⁵

Abstract

Prostate cancer (PCa) is one of the most common male malignancies. Serum prostate-specific antigen (PSA) is one of the most valuable biomarkers in tumor biology and remains the standard marker in detecting and monitoring PCa. However, the high number of serum PSA false positive and false negative results make the identification of novel biomarkers extremely welcome to improve our diagnostic accuracy in detecting PCa and distinguishing the aggressive from the indolent ones. In this study, we analyzed the current role of urinary gene fusion transcripts involving v-ets erythroblastosis virus E26 oncogene homolog, commonly known as ERG, and the androgen-regulated gene transmembrane protease, serine 2 (TMPRSS2), as a biomarker for PCa. Used as a single marker, urinary TMPRSS2:ERG has low sensitivity but high specificity. However, its combination with the other urinary marker PCa antigen 3 (PCA3) has been reported to provide high specificity and sensitivity. Finally, a commercially available assay combining serum PSA with urinary PCA3 and TMPRSS2:ERG provides a 90% specificity and 80% sensitivity in diagnosing PCa. Urinary TMPRSS2:ERG also seems to be indicative of PCa aggressiveness upon biopsy. Should these findings be confirmed in larger studies, urinary TMPRSS2:ERG might become a valuable test not only for diagnosing PCa but also for distinguishing the aggressive tumors from the indolent ones.

Clinical Genitourinary Cancer, Vol. 14, No. 2, 117-21 © 2016 Elsevier Inc. All rights reserved. **Keywords:** Diagnosis, Male genital neoplasm, Molecular biology, Prognosis, Urine marker

Introduction

Prostate cancer (PCa) is one of the most common male malignancies worldwide.¹ This finding could also be due to the extensive use of serum prostate-specific antigen (PSA) testing, which is currently the only widely used serum biomarker for PCa.² In the past 20 years, PSA testing has allowed clinicians to screen, diagnose earlier, and monitor PCa patients,³ ultimately leading to 2 apparently opposing facts: an increased cancer-specific mortality,⁴ and the diagnosis also of indolent tumors. This last fact, namely overdiagnosis/overtreatment of tumors not carrying a significant lethal potential, involves the drawbacks of unnecessary morbidity,⁵ and unnecessary health care costs.⁶ Another disadvantage of PSA testing is its low specificity (ranging from 25% to 40%). Increased serum PSA levels might be detected in several nonneoplastic diseases such as prostatitis and benign prostatic hyperplasia (BPH), thus representing false positive results (negative prostate biopsy in patients with PSA > 4 ng/mL); conversely, patients with normal (< 4 ng/mL) PSA levels might harbor PCa (false negative results).⁷ This led to the recent recommendation from the main US and European PCa guidelines against using PSA-based mass PCa screening.^{6,8} Because of these limitations of PSA testing, great efforts have been made to search for additional more specific biological markers (biomarkers), with a focus on PSA isoforms⁹ as well as the development of novel biomarkers.

A biomarker is defined as "a characteristic that is objectively measured and evaluated as an indicator of normal biological processes, pathogenic processes, or pharmacologic responses to a therapeutic intervention."¹⁰ Beyond its high diagnostic accuracy, an ideal biomarker in the clinical practice of PCa should be measurable with a simple, inexpensive, and repeatable test carried out in an easily available body fluid such as serum, urine, or prostatic fluid.¹¹

Because of the facility of collection and the fact that prostate cells are directly discharged into the urethra through prostatic ducts after applying pressure to the prostate, as is performed with a digital rectal

¹Department of Pathology, University of Foggia, Foggia, Italy

²Department of Biosciences, Biotechnologies, and Biopharmaceutics, University of Bari, Bari, Italy

³Department of Pathology and Diagnostic, University of Verona, Verona, Italy ⁴Anatomic Pathology, Card. G. Panico Hospital, Tricase, Italy ⁵Department of Urology and Renal Transplantation, University of Foggia, Foggia, Italy

Submitted: Aug 12, 2015; Revised: Nov 25, 2015; Accepted: Dec 9, 2015; Epub: Dec 17, 2015

Address for correspondence: Francesca Sanguedolce, MD, PhD, Via Masi, 7 - 71121 Foggia (FG), Italy

Fax: +390881736334; e-mail contact: fradolce@hotmail.com

TMPRSS2:ERG Fusion Transcript as a Biomarker

examination (DRE), urine has become the optimal substrate for noninvasive biomarker testing.¹²

Current technologies have widened the spectrum of putative biomarkers, by allowing the more and more careful analysis of DNA, mRNA, proteins, metabolites, or processes such as apoptosis, angiogenesis, or proliferation¹³ using new-generation methods such as DNA microarrays or quantitative and semiquantitative polymerase chain reaction (PCR)-based techniques.^{14,15}

Such technologies have allowed the identification of novel PCa biomarkers in tissue and, most importantly, in noninvasive samples.^{11,16,17} Nevertheless, no biomarker has so far replaced the routine use of PSA as a screening and follow-up method of PCa, because none have been through all 5 phases of testing (ie, preclinical exploratory studies, clinical assay and validation, retrospective longitudinal studies, prospective screening studies, and randomized controlled trials; reviewed in Huang et al¹⁸), which seal its official use in clinical practice.

Herein, we analyzed the current role of urinary transmembrane protease, serine 2 (TMPRSS2):v-ets erythroblastosis virus E26 oncogene homolog (ERG) as a biomarker for PCa.

Transmembrane Protease, Serine 2:ERG as a PCa Biomarker

On the basis of the compelling evidence that genomic rearrangements represent initial events in oncogenesis, gene fusion transcripts involving ERG (also known as TMPRSS2:ERG, or T2E), a member of the E 26 (ETS) family of oncogenestranscription factors in prostate cells, have been identified as promising urinary novel biomarkers.¹⁸ In 2005, using a new biostatistical method called cancer outlier profile analysis, Tomlins et al¹⁹ identified strong outlier profiles for the gene members of the ETS family of oncogenes in 16 PCa patients (55%), in keeping with the results of a previous study in which ERG overexpression was found in 59 (72%) PCa cases.²⁰ Moreover, by performing combined assays, they detected fusion of these ETS family members with the 5' untranslated region of the prostate-specific and androgen-regulated gene TMPRSS2 in 19 cases (>90%) overexpressing ERG or ets variant gene (ETV) 1, implying that the overexpression is most probably caused by the fusion.¹⁹ Of them, TMPRSS2 fusion with ERG is the main variant in approximately 50% (range, 40%-70%) of PCa patients, although other fusions involving the ETS family members ETV4 and ETV5 have been described as a rarer molecular event in PCa,^{21,22} which might account for the aberrant androgen-regulated overexpression of ETS family members in some PCa because TMPRSS2 is androgendependent.¹⁹ Because ERG proto-oncogene overexpression is the more commonly reported in PCa cells, TMPRSS2:ERG fusion is believed to play a main role in prostate tumorigenesis,²³ and has been recognized as the most frequent genetic anomaly recounted so far in human solid cancers.²⁴

From bench to bedside, such androgen-regulated gene fusion between TMPRSS2 and ERG might try to accomplish the task of detecting and managing PCa, because these TMPRSS2:ERG gene fusions were further described in almost 50% of the PSA-screened Caucasian PCa patients and, with lower incidence, in the Asian population.^{25,26} Such molecular markers are also seldom present in high-grade prostatic intraepithelial neoplasia (hg-PIN).¹⁹

A few years ago, Laxman et al detected the mRNA products of the TMPRSS2:ERG fusion gene in urine samples from PCa patients for the first time.²⁷ In a subsequent study, this urine test featured a quite low sensitivity of 37%, a specificity of 93%, and a positive predictive value of 94% in post-DRE urine samples from 78 men with PCa-positive biopsies and 30 men with PCa-negative biopsies using semiquantitative reverse transcription PCR (RT-PCR).²⁸

In another study on a small cohort of patients, no TMPRSS2:ERG fusion transcripts were found in urine samples obtained from women, healthy young men, and post-radical prostatectomy (RRP) patients¹⁷; however, TMPRSS2:ERG fusion transcripts were found in 34.8% of the urine samples from PCa patients compared with 18.2% from men with negative biopsies (which the authors suggest were false negative).

In a recent editorial article, Tomlins raised concern regarding the reliability of this test, because of the well known multifocality and heterogeneity of PCa,²⁹ thus suggesting that a dual outcome (positive/negative) of TMPRSS2:ERG expression in urine PCa patients, as used in some studies,²⁴ has intrinsic limitations so that it does not overcome the need for an optimal cutoff to maximize the sensitivity and specificity balance.²⁹

The TMPRSS2:ERG fusion has been also studied for its prognostic potential with controversial results. According to Leyten et al, the fact that cancer cells from high-risk PCa have a higher potential to infiltrate contiguous structures, resulting in their greater post-DRE release to the prostatic ducts, ultimately resulting in urine specimens bearing more TMPRSS2:ERG fusion mRNA, could account for the hypothesis that urine TMPRSS2:ERG might have prognostic value.¹⁶

The largest study so far, in a cohort of 1180 men treated with RRP with a median follow-up of 12.6 years, reported overexpression of ERG in 49% of tissue samples using immunohistochemistry, with a significant association with tumor stage, but not with Gleason score, metastases, biochemical recurrence, and canceroverall mortality,³⁰ and thus suggesting related that TMPRSS2:ERG is not a strong predictor of outcome in surgicallytreated PCa patients. Another study that assayed the urinary TMPRSS2:ERG fusion transcript in 37% of pre-biopsy PCa patients found no correlation was found with Gleason score assessed on biopsy, which can be a potential pitfall because of the usual occurrence of upgrading in RRP specimens.²⁸ Indeed, in a subsequent study that measured urine TMPRSS2:ERG transcript levels through a clinical-grade, transcription-mediated amplification assay, Tomlins et al showed a significant association with cancer volume and grade and upgrading of Gleason score at the time of RRP.³¹

There is a growing body of literature proposing that the presence of the TMPRSS2:ERG fusion is a possible prognosticator of PCa outcome. In a cohort of localized PCa patients treated by watchful waiting, TMPRSS2:ERG fusion was reported in association with Gleason score and cancer-specific death; the selection bias of this low-stage population possibly explains an incidence as low as 15% compared with the average 50% reported in most studies.³² However, in this cohort TMPRSS2:ERG fusion was detected in only 15% of the PCa patients compared with the average 50% reported in most studies, potentially because of the selection bias of this low-stage population.³² In keeping with such findings, Attard et al found that a deletion-based fusion detected using a fluorescent in situ hybridization assay on tissue microarray specimens was associated with clinical stage, Gleason score, baseline PSA, and a poor cancer-specific survival,³³ as confirmed by other studies.¹⁶

Ongoing research topics are the exploitment of TMPRSS2:ERG gene fusion in patients with localized PCa and hg-PIN deserving active surveillance. 34

Taken together, such studies highlight the limitations of TMPRSS2:ERG fusion assay as a urine biomarker of PCa for screening purposes; the main pitfalls are: (1) the lower incidence of the fusion in some cohorts of men (such as localized PCa patients) seem to provide lower screening sensitivity³²; (2) a suitable, comprehensive cutoff has not been identified yet, and the issues of PCa heterogeneity and multifocality seem to trouble its assessment; (3) the test estimates the quantity of TMPRSS2:ERG fusion RNA in urine samples in relation to PSA mRNA, to measure the amount of prostate cells in the urine sediment; the assay is considered invalid when the PSA level is too little³⁵; and (4) most studies describe a sensitivity that is too low to allow clinical value for the test alone, with a most recent systematic review and meta-analysis reporting a specificity of 86% and sensitivity of 45%.³⁶

Although the high specificity of TMPRSS2:ERG urine assay could be exploited for risk stratification of patients with a negative biopsy, thus urging an urgent rebiopsy or magnetic resonance imaging, this assay is not yet verified in a clinical setting as a PCa biomarker to point out the urge for rebiopsy.

In Table 1 the most relevant studies dealing with the role of TMPRSS2:ERG as a PCa biomarker are summarized.

V-ets Erythroblastosis Virus E26 Oncogene Homolog in a Panel of PCa Biomarkers

In keeping with the heterogeneity of the disease and the apparent low sensitivity of TMPRSS2:ERG, the construction of multiplexed models on the basis of a combination of cancer-specific biomarkers would be more appropriate to increase the overall diagnostic accuracy of this assay in the clinical management of PCa.³⁵

Francesca Sanguedolce et al

Biomarkers other than PSA have been developed recently. PCa antigen 3 (PCA3) is a prostate-specific noncoding mRNA that is raised in more than 95% of PCa, compared with benign prostate tissues,¹¹ thus being probed as a urine assay for PCa early detection³⁷ and also proved to be related to histologic grade and tumor volume in surgical specimens.³⁸ In the United States, Progensa PCA3 (Gen-Probe) is a commercially available assay used in the United States after the US Food and Drug Administration (FDA) approval in 2012 as a diagnostic test to help in risk stratification for patients with a negative prostate biopsy, to select those who will benefit from a repeat biopsy.

The combined approach of PCA3 and TMPRSS2:ERG gene fusion testing for PCa diagnosis is supported by the evidence of the different increase of each marker in prostatic diseases; in the recent in vitro study by Robert et al on 32 normal prostate tissues, 48 BPH, and 48 PCa,³⁹ PCA3 levels showed gradual increase from BPH to normal prostate tissue and PCa tissue (3 and 30 times, respectively) and the TMPRSS2:ERG gene fusion was detected in 4 (8.3%) of the BPH, 5 (15.6%) of the normal prostate tissue, and 24 (50%) of the PCa samples.³⁹ Although the authors showed that the use of both combined markers improved the sensitivity, because most false-negative results of the PCA3 test could be corrected using the TMPRSS2:ERG fusion assay, they also suggested the need to assess an optimal cutoff to withdraw all false positive results.³⁹

Interestingly, in one study that detected the TMPRSS2:ERG and PCA3 in urine samples from a huge cohort of 246 men,⁴⁰ it was reported that a significant lower area under the receiver operating characteristic curve (AUC) for TMPRSS2:ERG (0.63) than for PCA3 (0.74), and the use of combined parameters did not result in a significant increase in diagnostic accuracy compared with single markers.⁴⁰

However, through a series of reports over the past 7 years, the combined use of PCA3 and TMPRSS2:ERG gene fusion transcripts, also included in widely used PCa risk calculators, was shown to significantly increase the reliability at diagnosis of each single test, ^{16,28,31,37,41-43} and some authors claimed that they might be related to PCa aggressiveness. ^{31,38}

Table 1	Associa	ssociation Between Urine TMPRSS2:ERG Fusion and Clinicopathological Findings From Selected Studies				
Study		Sample Size	Study Design	Significant Correlation	Limitations	
DeMichelis et al ³²		111 Men with localized PCa	Population-based cohort study	PCa specific death	NA	
Tomlins et al ³¹		1312 Men	Cohort study	Clinically significant PCa at the time of biopsy and prostatectomy (tumor size, high Gleason score at the time of prostatectomy, upgrading of Gleason grade at the time of prostatectomy)	Most patients in the study (>85%) were Caucasian Men in this study were PSA-screened (and elected to undergo biopsy)	
Salami e	t al ⁴¹	45 Men	Cohort study	PCa found at biopsy The greatest discriminatory value in predicting PCa compared with PCA3 and PSA alone	Small sample size	
Cornu et	al ⁴⁵	291 Men	Cohort study	PCa at biopsy	Lack of a prospective design, thus the data are limited to odds ratios, which do not permit causality to be assessed	
Leyten et	t al ¹⁶	443 Men	Multicenter cohort study	Significant additional predictive value to the ERSPC risk calculator parameters to predict biopsy Gleason score and clinical tumor stage	Lack of correlation with Gleason score in radical prostatectomy specimens	

Abbreviations: ERG = v-ets erythroblastosis virus E26 oncogene homolog; ERSPC = European Randomised Study of Screening for Prostate Cancer; PCa = prostate cancer; PCA3 = prostate cancer antigen 3; PSA = prostate-specific antigen; TMPRSS2 = transmembrane protease, serine 2.

TMPRSS2:ERG Fusion Transcript as a Biomarker

In a recent review of the literature on the role of genetic analysis in PCa, the authors have pointed out PCA3 and TMPRSS2:ERG fusion transcripts as promising RNA diagnostic and predictive markers.⁴⁴

The first report by Hessels et al²⁸ was based on a prospective study on 108 prebiopsy post-DRE urine samples, 78 (72%) of which were later diagnosed with PCa upon biopsy. Because of its high specificity (93%), the combination of PCA3 and TMPRSS2:ERG increased the test sensitivity by 11% (from 62% for PCA3 alone to 73% for both markers). Such results have been subsequently corroborated by Cornu et al.⁴⁵ The latter showed that a PCA3 and TMPRSS2:ERG combined assay raised the outcome of the multivariate PCPT (PCa Prevention Trial) risk calculator in predicting cancer on biopsy by increasing the AUC from 0.66 to 0.75, although their findings were biased by the lack of uniformity of assay threshold among the centers involved in the study.

The diagnostic accuracy of the PCA3 and TMPRSS2:ERG transcripts assay has been studied also in combination with other markers. Laxman et al assayed the combination of 4 biomarkers, comprised of PCA3 and TMPRSS2:ERG, along with Golgi phosphoprotein 2 (GOLPH2) and serine protease inhibitor Kazal-type 1 (SPINK1), each one being an independent predictor of PCa, using a quantitative multiplexed RT-PCR analysis model; as a result, sensitivity and specificity for PCa detection in rebiopsies were up to 66% and 76%, respectively.⁴⁶ Subsequently, Cao et al reported a high diagnostic accuracy combined panel of PCA3, TMPRSS2:ERG, annexin A3, and sarcosine for PCa diagnosis (AUC 0.856 vs. PCA3: 0.739, TMPRSS2:ERG: 0.732, annexin A3: 0.728, and Sarcosine: 0.665).⁴⁷

In a more recent study, Salami et al⁴¹ supported the highest sensitivity (93%) of PCA3 and highest specificity (87%) of TMPRSS2:ERG fusion transcript for detection of PCa in a cohort of urine samples from 48 patients, 15 of whom were diagnosed with PCa. Multivariable algorithms including PCA3, urine TMPRSS2:ERG, and serum PSA, and PCA3 + TMPRSS2:ERG + PSA + DRE were used to diagnose PCa with high accuracy.^{41,48}

Furthermore, in a recent study, Leyten et al assessed the independent additional predictive value of urinary TMPRSS2:ERG to PCA3 and the ERSPC (European Randomised Study of Screening for Prostate Cancer) risk calculator parameters (serum PSA level, DRE abnormal/normal, transrectal ultrasound abnormal/normal, and prostate volume) in a cohort of 443 men comprised of 44% PCa patients. Their results showed a stepwise increase in AUC from 0.799 (the ERSPC risk calculator alone) to 0.833 (ERSPC + PCA3) to 0.842 (ERSPC + PCA3 + TMPRSS2); interestingly, TMPRSS2:ERG showed prognostic value comparable with PCA3.¹⁶ For the prediction of clinically significant PCa, TMPRSS2:ERG was more specific than PCA3 alone; the latter, along with TMPRSS2:ERG, raised its sensitivity to 88% not compromising its specificity.¹⁶ Despite all such evidence for the 2 RNA-based biomarkers, PCA3 and TMPRSS2:ERG, 49 the available studies did not address the antagonistic effect of PCA3 and ERG score in a multivariable model. Moreover, all the cited studies did not integrate testosterone serum levels or genetic analyses, although both have been shown to affect the detection of PCa as well as its biological behavior.⁵⁰

Having said this, there is already a commercially available diagnostic assay in the United States and Canada for PCa, with outcome as a percentage of risk, on the basis of blood PSA levels and urinary PCA3 and T2E test results, with a specificity and sensitivity as high as 90% and 80%, respectively,⁴¹ thus outperforming traditional population-based nomograms.¹⁸

Conclusions

Serum PSA remains one of the most useful biomarkers in cancer biology and is here to stay. However, because of its high number of false positive and false negative results, it requires experienced interpretation. Therefore, the identification of novel biomarkers that can be used alone or in combination with PSA is extremely welcome to improve our diagnostic accuracy in detecting PCa and distinguishing the aggressive from the indolent ones.

Because PCa cells discarded into the urethra after a DRE, urine represents a noninvasive, easy to obtain milieu to gain information on PCa-related markers. Nevertheless, so far only few markers have been validated in large cohort studies. Of them, only PCA3 and TMPRSS2:ERG fusion transcripts have been shown to be promising RNA markers for detecting PCa and predicting its aggressiveness. PCA3 has been recently introduced in clinical practice upon FDA approval, and combined assay of both has been marketed for clinical use having been shown to perform better than the single marker in detecting PCa.

A major challenge in PCa management is the risk stratification of patients eligible for delayed treatment because of localized disease. Even though further evidence is needed, there is ground for believing that urinary ERG might have a predictive value as well.

Some potential limitations of using urine, including dilution, variability of collection methods, confounding effects of other urinary components, and biomarker degradation, nevertheless do not impair the future exploitment of the diagnostic and predictive potentials of such novel assays.

Disclosure

The authors have stated that they have no conflicts of interest.

References

- 1. Siegel R, Naishadham D, Jemal A. Cancer statistics. CA Cancer J Clin 2013; 63: 11-30.
- Dijkstra S, Mulders PF, Schalken JA. Clinical use of novel urine and blood based prostate cancer biomarkers: a review. *Clin Biochem* 2014; 47:889-96.
- Bangma CH, van Schaik RH, Blijenberg BG, Roobol MJ, Lilja H, Stenman UH. On the use of prostate-specific antigen for screening of prostate cancer in European Randomised Study for Screening of Prostate Cancer. *Eur J Cancer* 2010; 46: 3109-19.
- Xia J, Gulati R, Au M, Gore JL, Lin DW, Etzioni R. Effects of screening on radical prostatectomy efficacy: the prostate cancer intervention versus observation trial. *J Natl Cancer Inst* 2013; 105:546-50.
- Gann PH, Fought A, Deaton R, Catalona WJ, Vonesh E. Risk factors for prostate cancer detection after a negative biopsy: a novel multivariable longitudinal approach. J Clin Oncol 2010; 28:1714-20.
- Moyer VA, on behalf of the U.S. Preventive Services Task Force. Screening for prostate cancer: U.S. Preventive Services Task Force Recommendation Statement. *Ann Intern Med* 2012; 157:120-34.
- Thompson IM, Pauler DK, Goodman PJ, et al. Prevalence of prostate cancer among men with a prostate-specific antigen level < or = 4.0 ng per milliliter. *N Engl J Med* 2004; 350:2239-46.
- Heidenreich A, Abrahamsson PA, Artibani W, et al. Early detection of prostate cancer: European Association of Urology recommendation. *Eur Urol* 2013; 64: 347-54.
- 9. Djavan B. Validity and legacy of prostate-specific antigen (PSA) and PSA-based parameters and isoforms in the new millennium. *Eur Urol* 2010; 57:928-9.

- Biomarkers Definitions Working Group. Biomarkers and surrogate endpoints: preferred definitions and conceptual framework. *Clin Pharmacol Ther* 2001; 69: 89-95.
- Hessels D, Klein Gunnewiek JM, van Oort I, et al. DD3(PCA3)-based molecular urine analysis for the diagnosis of prostate cancer. *Eur Urol* 2003; 44:8-15, discussion 15–16.
- 12. Truong M, Yang B, Jarrard D. Towards the detection of prostate cancer in urine: a critical analysis. J Urol 2013; 189:422-9.
- 13. Hayes DF, Bast RC, Desch CE, et al. Tumor marker utility grading system: a framework to evaluate clinical utility of tumor markers. *J Natl Cancer Inst* 1996; 88:1456-66.
- Lapointe J, Li C, Higgins JP, et al. Gene expression profiling identifies clinically relevant subtypes of prostate cancer. *Proc Natl Acad Sci U S A* 2004; 101:811-6.
- Taylor BS, Schultz N, Hieronymus H, et al. Integrative genomic profiling of human prostate cancer. *Cancer Cell* 2010; 18:11-22.
- Leyten GH, Hessels D, Jannink SA, et al. Prospective multicentre evaluation of PCA3 and TMPRSS2-ERG gene fusions as diagnostic and prognostic urinary biomarkers for prostate cancer. *Eur Urol* 2014; 65:534-42.
- Nguyen PN, Violette P, Chan S, et al. A panel of TMPRSS2:ERG fusion transcript markers for urine-based prostate cancer detection with high specificity and sensitivity. *Eur Urol* 2011; 59:407-14.
- Huang JG, Campbell N, Goldenberg SL. PSA and beyond: biomarkers in prostate cancer. *BCMJ* 2014; 56:334-41.
- Tomlins SA, Rhodes DR, Perner S, et al. Recurrent fusion of TMPRSS2 and ETS transcription factor genes in prostate cancer. *Science* 2005; 310:644-8.
- Petrovics G, Liu A, Shaheduzzaman S, et al. Frequent overexpression of ETSrelated gene-1 (ERG1) in prostate cancer transcriptome. *Oncogene* 2005; 24: 3847-52.
- 21. Tomlins SA, Mehra R, Rhodes DR, et al. TMPRSS2:ETV4 gene fusions define a third molecular subtype of prostate cancer. *Cancer Res* 2006; 66:3396-400.
- Helgeson BE, Tomlins SA, Shah N, et al. Characterization of TMPRSS2:ETV5 and SLC45A3:ETV5 gene fusions in prostate cancer. *Cancer Res* 2008; 68:73-80.
- 23. Prensner JR, Chinnaiyan AM. Oncogenic gene fusions in epithelial carcinomas. Curr Opin Genet Dev 2009; 19:82-91.
- Perner S, Mosquera JM, Demichelis F, et al. TMPRSS2-ERG fusion prostate cancer: an early molecular event associated with invasion. *Am J Surg Pathol* 2007; 31:882-8.
- Magi-Galluzzi C, Tsusuki T, Elson P, et al. TMPRSS2–ERG gene fusion prevalence and class are significantly different in prostate cancer of Caucasian, African-American and Japanese patients. *Prostate* 2011; 71:489-97.
- Tomlins SA, Bjartell A, Chinnaiyan AM, et al. ETS gene fusions in prostate cancer: from discovery to daily clinical practice. *Eur Urol* 2009; 56:275-86.
- Laxman B, Tomlins SA, Mehra R, et al. Noninvasive detection of TMPRSS2:ERG fusion transcripts in the urine of men with prostate cancer. *Neoplasia* 2006; 8: 885-8.
- 28. Hessels D, Smit FP, Verhaegh GW, et al. Detection of TMPRSS2-ERG fusion transcripts and prostate cancer antigen 3 in urinary sediments may improve diagnosis of prostate cancer. *Clin Cancer Res* 2007; 13:5103-8.
- 29. Tomlins SA. Urine PCA3 and TMPRSS2:ERG using cancer-specific markers to detect cancer. *Eur Urol* 2014; 65:543-5.
- Pettersson A, Graff RE, Bauer SR, et al. The TMPRSS2:ERG rearrangement, ERG expression, and prostate cancer outcomes: a cohort study and meta-analysis. *Cancer Epidemiol Biomarkers Prev* 2012; 21:1497-509.
- Tomlins SA, Aubin SM, Siddiqui J, et al. Urine TMPRSS2:ERG fusion transcript stratifies prostate cancer risk in men with elevated serum PSA. *Sci Transl Med* 2011; 3:94ra72.

- 32. Demichelis F, Fall K, Perner S, et al. TMPRSS2:ERG gene fusion associated
- with lethal prostate cancer in a watchful waiting cohort. Oncogene 2007; 26: 4596-9.
- 33. Attard G, Clark J, Ambroisine L, et al. Duplication of the fusion of TMPRSS2 to ERG sequences identifies fatal human prostate cancer. *Oncogene* 2008; 27: 253-63.
- 34. Park K, Dalton JT, Narayanan R, et al. TMPRSS2:ERG gene fusion predicts subsequent detection of prostate cancer in patients with high-grade prostatic intraepithelial neoplasia. J Clin Oncol 2014; 32:206-11.
- Prensner JR, Rubin MA, Wei JT, Chinnaiyan AM. Beyond PSA: the next generation of prostate cancer biomarkers. *Sci Transl Med* 2012; 4:127rv3.
- Yao Y, Wang H, Li B, Tang Y. Evaluation of the TMPRSS2:ERG fusion for the detection of prostate cancer: a systematic review and meta-analysis. *Tumour Biol* 2014; 35:2157-66.
- Deras IL, Aubin SM, Blase A, et al. PCA3: a molecular urine assay for predicting prostate biopsy outcome. J Urol 2008; 179:1587-92.
- Ploussard G, Durand X, Xylinas E, et al. Prostate cancer antigen 3 score accurately predicts tumour volume and might help in selecting prostate cancer patients for active surveillance. *Eur Urol* 2011; 59:422-9.
- **39.** Robert G, Jannink S, Smit F, et al. Rational basis for the combination of PCA3 and TMPRSS2:ERG gene fusion for prostate cancer diagnosis. *Prostate* 2013; 73: 113-20.
- 40. Stephan C, Jung K, Semjonow A, et al. Comparative assessment of urinary prostate cancer antigen 3 and TMPRSS2:ERG gene fusion with the serum [-2]proprostate-specific antigen-based prostate health index for detection of prostate cancer. *Clin Chem* 2013; 59:280-8.
- Salami SS, Schmidt F, Laxman B, et al. Combining urinary detection of TMPRSS2:ERG and PCA3 with serum PSA to predict diagnosis of prostate cancer. Urol Oncol 2013; 31:566-71.
- Ficarra V, Novara G, Zattoni F. The role of the prostate cancer antigen 3 (PCA3) test for the diagnosis of prostate cancer in the era of opportunistic prostate-specific antigen screening. *Eur Urol* 2010; 58:482-4.
- Auprich M, Haese A, Walz J, et al. External validation of urinary PCA3-based nomograms to individually predict prostate biopsy outcome. *Eur Urol* 2010; 58: 727-32.
- 44. Alvarez-Cubero MJ, Saiz M, Martinez-Gonzalez LJ, Alvarez JC, Lorente JA, Cozar JM. Genetic analysis of the principal genes related to prostate cancer: a review. *Urol Oncol* 2013; 31:1419-29.
- 45. Cornu JN, Cancel-Tassin G, Egrot C, Gaffory C, Haab F, Cussenot O. Urine TMPRSS2:ERG fusion transcript integrated with PCA3 score, genotyping, and biological features are correlated to the results of prostatic biopsies in men at risk of prostate cancer. *Prostate* 2013; 73:242-9.
- **46.** Laxman B, Morris DS, Yu J, et al. A first-generation multiplex biomarker analysis of urine for the early detection of prostate cancer. *Cancer Res* 2008; 68: 645-9.
- Cao DL, Ye DW, Zhang HL, Zhu Y, Wang YX, Yao XD. A multiplex model of combining gene-based, protein-based, and metabolite-based with positive and negative markers in urine for the early diagnosis of prostate cancer. *Prostate* 2011; 71:700-10.
- Dimitriadis E, Kalogeropoulos T, Velaeti S, et al. Study of genetic and epigenetic alterations in urine samples as diagnostic markers for prostate cancer. *Anticancer Res* 2013; 33:191-7.
- Haese A, de la Taille A, van Poppel H, et al. Clinical utility of the PCA3 urine assay in European men scheduled for repeat biopsy. *Eur Urol* 2008; 54:1081-8.
- Cornu JN, Drouin S, Cancel-Tassin G, et al. Impact of genotyping on outcome of prostatic biopsies: a multicenter prospective study. *Mol Med* 2011; 17:473-7.